

This article was downloaded by:

On: 25 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597286>

Inhibition of HIV-1 Replication in Macrophages by Red Blood Cell-Mediated Delivery of a Heterodinucleotide of Lamivudine and Tenofovir

P. Franchetti^a; L. Cappellacci^a; R. Petrelli^a; P. Vita^a; M. Grifantini^a; L. Rossi^b; F. Pierigé^b; S. Serafini^b; M. Magnani^b; E. Balestra^c; C. -F. Perno^c

^a Department of Chemical Sciences, University of Camerino, Camerino, Italy ^b Institute of Biochemistry "G. Fornaini," University of Urbino, Urbino, Italy ^c Department of Experimental Medicine, University of Rome "Tor Vergata," Rome, Italy

To cite this Article Franchetti, P. , Cappellacci, L. , Petrelli, R. , Vita, P. , Grifantini, M. , Rossi, L. , Pierigé, F. , Serafini, S. , Magnani, M. , Balestra, E. and Perno, C. -F.(2007) 'Inhibition of HIV-1 Replication in Macrophages by Red Blood Cell-Mediated Delivery of a Heterodinucleotide of Lamivudine and Tenofovir', *Nucleosides, Nucleotides and Nucleic Acids*, 26: 8, 953 — 957

To link to this Article: DOI: 10.1080/15257770701508067

URL: <http://dx.doi.org/10.1080/15257770701508067>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

INHIBITION OF HIV-1 REPLICATION IN MACROPHAGES BY RED BLOOD CELL-MEDIATED DELIVERY OF A HETERODINUCLEOTIDE OF LAMIVUDINE AND TENOFOVIR

P. Franchetti, L. Cappellacci, R. Petrelli, P. Vita, and M. Grifantini □

Department of Chemical Sciences, University of Camerino, Camerino, Italy

L. Rossi, F. Pierigé, S. Serafini, and M. Magnani □ *Institute of Biochemistry “G. Fornaini,” University of Urbino, Urbino, Italy*

E. Balestra and C.-F. Perno □ *Department of Experimental Medicine, University of Rome “Tor Vergata,” Rome, Italy*

□ *Homo- and heterodimers of nucleoside/nucleotide analogues as reverse transcriptase inhibitors are effective on HIV-1-infected human monocyte-derived macrophages (M/M) compared to the single drugs or their combination. Since the combined treatment of lamivudine (3TC) and tenofovir ((R)PMPA) has an antiretroviral efficacy and a synergic effect respect to separate drugs, the heterodinucleotide 3TCpPMPA was synthesized. A single administration of the dimer as free drug or 3TCpPMPA-loaded RBC selectively targeted to M/M was able to almost completely protect macrophages from “de novo” infection.*

Keywords Heterodinucleotides; anti-HIV-1 activity; human macrophages

INTRODUCTION

The combination of nucleoside analogues as reverse transcriptase inhibitors (NRTIs) with nucleotide analogues or other antiviral drugs resulted in effective therapy for the treatment of human viral infections. Despite the clinical benefit associated with the highly active antiretroviral therapy (HAART), current drugs are not able to eradicate HIV-1 simply due to the persistence of the virus in cellular reservoirs, predominantly long-lived memory CD4⁺ T cells and cells of the macrophage lineage.^[1] Indeed, HIV-1 has been recovered from the peripheral blood monocytes of patients with maximal viral suppression while on HAART.^[2] Viral reservoirs established

This work was supported by grants from AIDS Project of the Italian National Institute of Health and by FIRB funds. (Red blood cells as drug carriers, RBNE01TBTR.)

Address correspondence to Mario Grifantini, Department of Chemical Sciences, University of Camerino, 62032 Camerino, Italy. E-mail: mario.grifantini@unicam.it

early during the infection remain unaffected by antiretroviral therapy for a long time and are able to replenish systemic infection following the failure or cessation of the treatment.

Since the crucial role of the monocyte-derived macrophages (M/M) as a HIV-1 reservoir, therapeutic strategy must consider the inhibition of the viral replication in M/M.^[3]

However, since macrophages are resting cells, they phosphorylate (activate) the antiviral nucleoside analogues at a rate lower than that of peripheral blood mononuclear cells.^[4] Thus, the delivery of phosphorylated nucleoside analogues directly into infected-macrophages could result in highly anti-HIV-1 activity.

On this basis, some antiviral double-drugs (homo- and heterodinucleotides) as reverse transcriptase inhibitors and as potential pro-drugs with efficacy against HIV-1-infected M/M have been reported by us.^[5] Herein, we report on the synthesis of a new heterodinucleotide 3TCpPMPA consisting of the potent anti-HIV-1 nucleoside and nucleotide analogues lamivudine (3TC) and tenofovir ((*R*)PMPA), respectively.

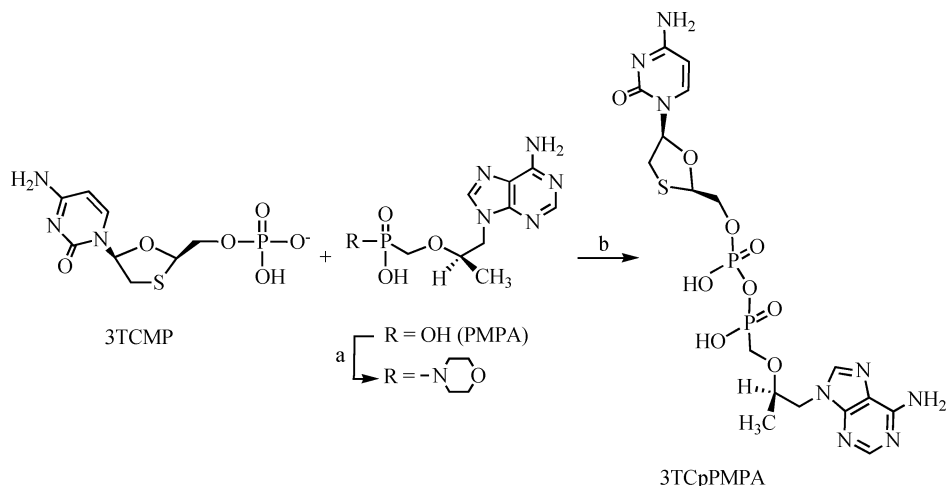
The combination of tenofovir and lamivudine has already been suggested in therapy guidelines for HAART-naïve patients.^[6] The efficacy of the dimer 3TCpPMPA to protect the refractory cell compartment against de novo HIV infection by simultaneous administration of 3TC and (*R*)PMPA in a single molecule has been assessed.

RESULTS AND DISCUSSION

The heterodinucleotide consisting of 3TC and (*R*)PMPA bound together by a phosphate bridge was synthesized by coupling the mono *n*-tri-butylammonium salt of β -L-3'-thia-2',3'-dideoxy-cytidine monophosphate (3TCMP) with (*R*)-9-(2-phosphonometoxypyl)adenine as a morpholidate derivative (Scheme 1).

At first, the metabolism of 3TCpPMPA in plasma obtained from healthy seronegative donors was determined. As shown in Figure 1, human plasma possesses enzymes able to cleave the phosphate bridge of 3TCpPMPA with the stoichiometric production of PMPA and 3TC (the latter is quickly obtained starting from 3TCMP). The results obtained show that 3TCpPMPA is stable enough in human plasma ($t_{1/2} = 4$ hours) and suggest the possibility of using 3TCpPMPA as a prodrug of 3TC and PMPA.

The ability of the heterodimer to protect the infected human macrophages against de novo HIV-1 infection also was determined. Compound was administered to macrophages and maintained all through experiment at a concentration of 1.0 μ M and virus production was evaluated over a period of 35 days. The results show an almost 100% inhibition of viral replication (Table 1).



SCHEME 1 Reagents and conditions: a) morpholine, dicyclohexylcarbodiimide, *t*-BuOH/H₂O, reflux; b) N,N-dimethylformamide, 40°C.

The selective delivery of 3TCpMPA to macrophages was performed using a drug targeting system. This system is based on the use of autologous artificially-aged red blood cells (RBC)⁷ that allows the direct administration of nucleoside analogues, already in phosphorylated form, to macrophages. The heterodinucleotide was encapsulated into autologous erythrocytes by

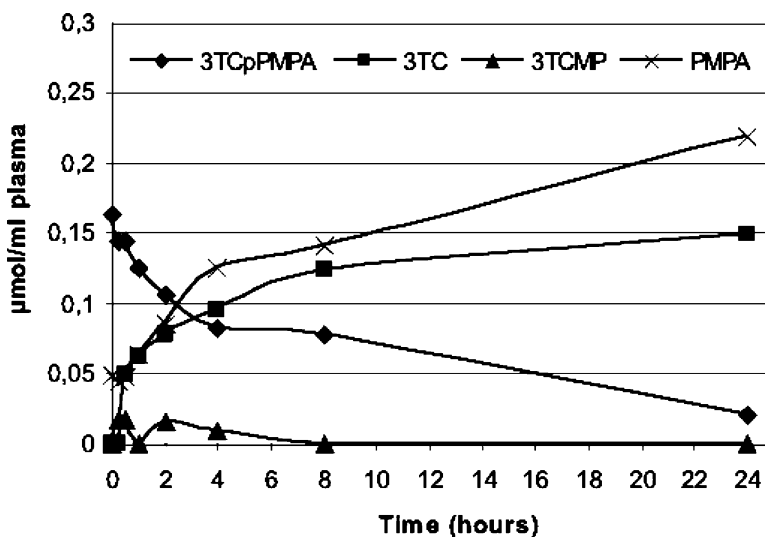


FIGURE 1 Metabolism of 3TCpMPA in human plasma. 3TCpMPA (0.25 mM) was incubated in human plasma for 24 hours at 37°C. Perchloric acid extracts were prepared at different incubation times and analyzed by HPLC, as described above. All values are the mean (\pm S.D.) of three different experiments.

TABLE 1 Percentage of virus production in HIV-1-infected human macrophages

Virus production (% of infected control)	
Time (days)	3TCpPMPA (1.0 μ M)
14	0
21	0
28	0.3
35	0.5

Values are expressed in percentage of p24 production respect to untreated infected macrophages. All values are the mean (\pm S.D.) of sextuplicate cultures of three representative experiments and show the percentage of inhibition in p24 produced until 35 days after infection. One hundred percent virus production corresponds to 23625 ± 2902 pg p24 antigen/mL.

a procedure of hypotonic dialysis, isotonic resealing, and reannealing.^[7] 3TCpPMPA-loaded erythrocytes were modified to increase their phagocytosis by human macrophages. Then, the cells were infected by HIV-1_{Ba-L} and inhibition of HIV-1 replication was assessed by HIV p24^{gag} quantification. When 3TCpPMPA was selectively targeted to the macrophagic compartment by a single administration of loaded erythrocytes, the protection of macrophages from de novo infection was nearly complete (99% protection 3 weeks posttreatment).

In conclusion, the heterodinucleotide 3TCpPMPA could act as an efficient prodrug that, by the conversion in 3TCMP and PMPA inside macrophages, is able to protect the refractory cell compartment against de novo HIV-1 infection for a long time.

REFERENCES

1. Marcello, A. Latency: The hidden HIV-1 challenge. *Retrovirology* **2006**, 3, 7.
2. a) Crowe, S.M.; Sonza, S. HIV-1 can be recovered from a variety of cells including peripheral blood monocytes of patients receiving highly active antiretroviral therapy: a further obstacle to eradication. *J. Leukoc. Biol.* **2000**, 68, 345–350; b) Zhu, T.; Muthui, D.; Holte, S.; Nickle, D.; Feng, F.; Brodie, S.; Hwangbo, Y.; Mullins, J.L.; Corey, L. Evidence for human immunodeficiency virus type 1 replication *in vivo* in CD14(+) monocytes and its potential role as a source of virus in patients on highly active antiretroviral therapy. *J. Virol.* **2002**, 76, 707–716; c) Sonza, S.; Mutimer, H.P.; Oelrichs, R.; Jardine D.; Harvey K.; Dunne, A.; Purcell, D.F.; Birch, C.; Crowe, S.M. Monocytes harbour replication-competent, non-latent HIV-1 in patients on highly active antiretroviral therapy. *AIDS* **2001**, 15, 17–22.
3. Acquaro, S.; Svicher, V.; Schols, D.; Pollicita, M.; Antinori, A.; Balzarini, J.; Perno, C.-F. Mechanisms underlying activity of antiretroviral drugs in HIV-1-infected macrophages: New therapeutic strategies. *J. Leukoc. Biol.* **2006**, 80, 1103–1110.
4. Perno, C.-F.; Yarchoan, R.; Cooney, D.A.; Hartman, N.R.; Gartner, S.; Popovic, M.; Hao, Z.; Gerrard, T. L.; Wilson, Y.A.; Johns, D.G. Inhibition of human immunodeficiency virus (HIV-1/HTLV-III_{Ba-L}) replication in fresh and cultured human peripheral blood monocytes/macrophages by azidothymidine and related 2',3'-dideoxynucleosides. *J. Exp. Med.* **1988**, 168, 1111–1125.

5. Rossi, L.; Serafini, S.; Franchetti, P.; Cappellacci, L.; Fraternali, A.; Casabianca, A.; Brandi, G.; Pierigé, F.; Perno, C.-F.; Balestra, E.; Benatti, U.; Millo, E.; Grifantini, M.; Magnani, M. Targeting nucleotide dimers containing antiviral nucleosides. *Curr. Med. Chem. Anti-Infective Agents* **2005**, *4*, 37–54 and references cited therein.
6. AIDSinfo. Guidelines for the use of antiretroviral agents in HIV-1-infected adults and adolescents. Rockville: A Service of the U.S. Department of Health and Human Services; 6 October 2005. <http://aidsinfo.nih.gov/ContentFiles/AdultandAdolescentGL.pdf>. Accessed August 2, 2006.
7. Magnani, M.; Rossi, L.; Brandi, G.; Schiavano, G.F.; Montroni, M.; Piedimonte, G. Targeting antiretroviral nucleotide analogues in phosphorylated form to macrophages: *in vitro* and *in vivo* studies. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 6477–6481.